

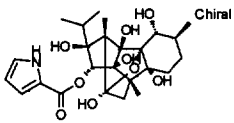
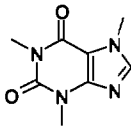
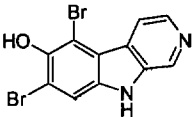
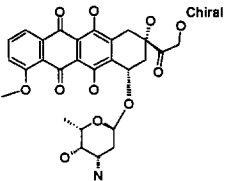
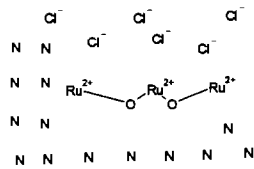
| | | | | | |
|-----------------------|-------------------|--|------------|------------|------------|
| Ryanoids | Ryanodine |  | Inactive | Antagonist | Antagonist |
| Xanthines | Caffeine |  | Inactive | Agonist | Agonist |
| Eudostomins | Bromoeudistomin D |  | Not tested | Agonist | Not tested |
| Antraquinones | Doxorubicin |  | Not tested | Inactive | Inactive |
| Polycationic reagents | Ruthenium Red |  | Displacer | Inactive | Antagonist |

Table 7

5

Data For Selected Anthranilamide Analogs (as shown in Figure 2)
Tested in Radioligand Displacement and Calcium Release Assays

| Molecule | R1 | R2 | R3 | R4 | R5 | Assay Type | | |
|----------|-----------|----|-----|-----|----|---------------------------------------|-----------------------------------|---------------------|
| | | | | | | Displacement IC ₅₀ (nM) | Calcium Release Threshold (nM) | |
| | | | | | | | <i>P. americana</i> | <i>P. americana</i> |
| Group 1 | | | | | | | | |
| Cpd 1 | isopropyl | H | CH3 | CF3 | F | < 300 | < 1000 | < 300 |
| Cpd 2 | isopropyl | H | CH3 | CF3 | Cl | < 300 | < 300 | < 300 |
| Cpd 3 | isobutyl | Br | CH3 | CF3 | Cl | < 300 | Not tested | Not tested |
| Cpd 4 | isopropyl | Br | CH3 | CF3 | F | Not tested | Not tested | Not tested |
| Group 2 | | | | | | | | |
| Cpd 5 | isopropyl | H | CH3 | CF3 | Cl | < 300 | < 300 | < 300 |

| | | | | | | | | |
|----------------|---------------------------------|----|-----------------|-----------------|-----------------|------------|------------|------------|
| Cpd 6 | CH ₃ | H | CH ₃ | CF ₃ | Cl | < 300 | < 300 | Not tested |
| Cpd 7 | isopropyl | H | CH ₃ | Br | Cl | < 300 | < 300 | Not tested |
| Cpd 8 | isopropyl | H | CH ₃ | Cl | Cl | < 300 | < 300 | Not tested |
| Cpd 9 | isopropyl | Br | CH ₃ | CF ₃ | Cl | < 300 | < 300 | Not tested |
| Cpd 10 | isopropyl | Cl | CH ₃ | CF ₃ | Cl | < 300 | < 300 | Not tested |
| Cpd 11 | CH ₃ | Cl | CH ₃ | CF ₃ | Cl | < 300 | < 300 | < 300 |
| Cpd 12 | isopropyl | Br | CH ₃ | Br | Cl | < 300 | < 300 | Not tested |
| Cpd 13 | isopropyl | Cl | CH ₃ | Br | Cl | < 300 | Not tested | Not tested |
| Cpd 14 | CH ₃ | Cl | CH ₃ | Br | Cl | < 300 | < 300 | < 300 |
| Cpd 15 | CH ₃ | Br | CH ₃ | CF ₃ | Cl | < 300 | < 300 | Not tested |
| Cpd 16 | isopropyl | Cl | CH ₃ | Cl | Cl | < 300 | < 300 | < 300 |
| Cpd 17 | CH ₃ | Cl | CH ₃ | Cl | Cl | < 300 | < 300 | Not tested |
| Cpd 18 | CH ₂ CH ₃ | Br | Br | CF ₃ | Cl | < 300 | Not tested | Not tested |
| Group 3 | | | | | | | | |
| Cpd 19 | isopropyl | H | Cl | CF ₃ | CH ₃ | < 3000 | < 10000 | < 10000 |
| Cpd 20 | isobutyl | H | Cl | CF ₃ | CH ₃ | Not tested | Not tested | < 3000 |
| Cpd 21 | isopropyl | H | CH ₃ | CF ₃ | CH ₃ | Not tested | Not tested | < 3000 |

EXAMPLE 10

Roach Muscle Membrane Preparation

The procedure for preparation of membranes from cockroach femoral muscle for radioligand binding studies was essentially as described by Schmitt, et.al. (1996) *Pesticide Science* 48:375-385. A brief description of the steps involved is as follows:

Mid and hind legs from 6-9 week old cockroaches were excised and immediately placed in liquid nitrogen. Excised legs could be stored at -80 °C for several months, if necessary, prior to further fractionation. Typically, membrane preparations started with approximately 55 g (wet weight) of excised legs. All subsequent steps were carried out at 4°C.

50 mM Tris-HCL, pH 7.4 was added at a ratio of 9 ml buffer to 1 g (wet weight) of legs. The legs were homogenized on ice using a Polytron (Brinkmann Instruments, Westbury, NY) for 2 min at Setting #3. In order to remove all fragments of cuticle, the homogenate was first passed through two layers of cheesecloth and then through one layer of glass wool overlaid on two layers of cheesecloth. On occasion, the cheesecloth clogged with particulates prior to passing all material through. In such cases, fresh cheesecloth (and glass wool) was used for the remainder of the suspension. The filtrate was collected in 50 ml centrifuge tubes and centrifuged in a Sorvall SS-34 rotor at 1,950 x g_{max} for 20 min. The supernatant was collected and centrifuged for 30 min at 30,000 x g_{max}. The